Co-selection of cognate antibody–antigen pairs by selectively-infective phages

Claus Krebber**, Stefania Spada**, Dominique Desplanq***, Andreas Plückthun*

Biochemisches Institut, Universität Zürich, Winterthurerstr. 190, CH-8057 Zürich, Switzerland

Received 9 November 1995

Abstract We have developed a chloramphenicol resistant derivative of fd phage with which cognate pairs of antibodies and antigens can be selected. The phage genome encodes a fusion of single-chain antibody to the C-terminal domain of gIIIP, rendering the phage non-infective. The antigen fused to the N-terminal domains of gIIIP is encoded in the same phage genome. Antigen and antibody fusion interact with each other in the periplasm of the phage-producing cell, restoring infectivity. This system has a very low background and will allow simultaneous randomisation of antibody and antigen.

Key words: Selectively-infective phage; Phage display; Antibody engineering; Filamentous phage; Single-chain Fv fragment

1. Introduction

Phage display technology has been successfully used to select individual peptide or protein sequences that bind a given target from libraries of filamentous phages [1,2]. Applications of this technology have included determination of the peptide epitope of an antibody [3] and selection for antibody fragments that bind to peptidic antigens [4]. We have now devised a selectively-infective phage (SIP) in which cognate pairs of peptidic antigen and antibody can be selected at the same time.

In traditional phage panning, libraries of the peptide or protein of interest are fused to an intact gene III phage protein (gIIIP) [5] or to its C-terminal domain requiring the use of helper phages to supply wt gIIIP [6]. The resulting display-phages are adsorbed to the binding partner and thus, either directly or indirectly, to a solid support. The major technical challenge is to separate phages bound by the cognate interaction from the great majority that are bound by nonspecific interactions. This is usually achieved by means of multiple rounds of binding and subsequent reinfection. The nature of these experiments allows the randomisation of only one of the reaction partners.

In our system, the antibody-displaying phage is itself non-infectious and acquires the ability to infect cells only upon the occurrence of a cognate interaction between antibody and antigen in the periplasm, directly liberating infectious phage from the host (Fig. 1). This selective infectivity is achieved by exploiting the modular structure of gIIIP, a protein of three domains of 68, 131 and 150 amino acids separated by glycine-rich linkers of 18 and 39 amino acids, respectively (Fig. 2) [7]. The first two domains (N1-N2) have been shown to be responsible for the docking of the phage to the F-pili of male E. coli and subsequent penetration of the bacterial membrane [8,9,10], while the C-terminal domain (CT) is presumed to have a structural role, making contact with the DNA and forming the tip of the phage particle [11].

It has previously been reported that phagemids can be used to display one of the interaction partners on the phage by fusing it to the N-terminus of the third domain of gIIIP (CT). Infection can then be restored by the addition of the first domain (N1) in vitro [12], or of the first two domains (N1-N2) in vivo [13], fused to the cognate partner. To package such phagemids a second genetic entity, a helper phage, that provides the genes for all phage functions except the wt gIIIP (dgIIIP), is needed [14,15].

In order to overcome the problems intrinsic to these dgIIIP (see below), we have engineered a single genetic package, a phage, which encodes both an antibody-CT fusion (scFv-CT) and a N1-N2-antigen fusion (N1-N2-Ag). This allows the formation of antibody–antigen complexes in the phage-producing cell, in the absence of any external factor, thus restoring phage infectivity. We have now developed a very stable phage vector system which does not undergo noticeable genetic rearrangement. We have therefore devised an extremely simple system, relying on the transmission of antibiotic resistance by phage infection of bacteria, for delineating pairs of interacting molecules in vivo.

2. Materials and methods

2.1. Bacteria, phages, plasmids

XL1-Blue (Stratagene) (recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac [F' proAB lacPZAM15 Tn10 (Tet')]) was used as donor. XL1-Blue MRF' Kan (Stratagene) (A(mcrA)183 A(mcrCB-hsdSMRmrr)173 endA1 supE44 thi-1 relA1 gyrA96 relA1 lac [F' proAB lacPZAM15 Tn5 (Kan')]) was used as recipient. fd phage was obtained from American Type Culture Collection (ATCC: 15669-B2). Resistance genes were PCR-amplified from pACYC174 (cm, p), pACYC188 (kan') and pUC19 (amp').

2.2. scFv gene synthesis

The single-chain Fv fragment (scFv) in the orientation V\textsubscript{L}-linker-V\textsubscript{H}
of the antibody 17/9 [16] was obtained by gene synthesis [17]. The V<sub>i</sub> domain carries a three amino acid long FLAG [18]. To eliminate any oligomerisation tendency of the scFv, we have used a 30 amino acid linker between the V<sub>h</sub> and V<sub>i</sub> domains [19].

2.3. ELISA

The hap-peptide KNSPYDVPDYASLRS was synthesised on an ABI 430A Peptide Synthesizer (Applied Biosystems Inc.) using the FastMoc strategy [20]. The peptide was subsequently coupled to denatured and reduced transferrin by the heterobifunctional linker 3-maleimidobenzoyl-N-hydroxy-succinimide (Fluka) [21]. This was coated at 20 μg per well onto a Maxisorp ELISA plate (Nunc) at 4°C overnight. After 1 h blocking with 5% skim milk in PBS phage dilutions in PBS were incubated in the wells for 2 h at 26°C. The bound phages were detected with 1:5000 sheep anti-M13 IgG conjugated to horseradish peroxidase (Pharmacia) with soluble substrate (Boehringer).

2.4. Phage construction

Cloning was carried out according to standard methods [22]. PCR was done with Vent-Polymerase (New England Biolabs) according to the manufacturer.

2.5. Selective infection experiments

An overnight culture of XL1-Blue, containing phage DNA, was used to inoculate 2 x YT medium, supplemented with 1% glucose and antibiotics (34 μg/ml cam, 5 μg/ml tet). Cells were grown at 37°C until an OD<sub>600</sub> of 0.5 was reached. After centrifugation the pellet was resuspended in 2 x YT medium containing the appropriate antibiotics and 0.5 mM IPTG, to induce the expression of the two fusion proteins under the control of the lac promoters (lac<sup>+</sup>), allowing packaging of the phages. After overnight growth at 26°C, phage particles were harvested from the culture supernatant by precipitation with polyethylene glycol and centrifugation. To quantify infectivity, phages were incubated with 1 ml of XL1-Blue MR<sup>F</sup> Kan (OD<sub>600</sub> = 0.5) for 1 h at 37°C with gentle agitation. Bacteria were plated on 2 x YT-agar containing 50 μg/ml kan, 34 μg/ml cam, 1% glucose. Phage clones were analysed either by restriction analysis of miniprep DNA or growth of the colonies in microtiter wells followed by ELISA testing of the phage-containing supernatants for antigen binding.

3. Results and discussion

Phage particles can in principle be produced by a phagemid/helper phage system. In order to obtain a phage which is infective only when a cognate interaction between antigen and antibody takes place (SIP), the helper phage is not allowed to deliver any wt gIII to the phage particles (gIII HP). This gIII HP must be able to infect a phagemid library, however, for which it needs gIII protein. This special helper phage must therefore be produced in a cell which harbours an unpackageable gIII<sub>lp</sub> expression plasmid [12,15]. In attempting to devise such a system, we found that all gIII<sub>lp</sub>expression plasmids tested, derived from pUC, pACYC or pBR vectors, were packaged with low efficiencies due to the presence of unidentified cryptic packaging signals (data not shown). This led to the production of wt gIII<sub>lp</sub> in the pool of display phages, rendering these infectious and thus losing the 'non-infectivity' which is the prerequisite for the selectivity of the infection process. This may also explain the background previously observed by other workers [12,15]. Alternatively, the phagemid library can be transformed into gIII HP-containing cells [13]. Use of fCA55 and fKN16 [14] gives rise, however to the assembly of 'stump' CT [14], competing with the display of antibody fusion on the phage. This will lead to varying, but always low, antibody display rates that are dependent on the differing folding efficiencies of scFvs [24]. Furthermore, in our experience gIII M13KO7, from which gIII is precisely deleted, showed high genetic instability and lowered the viability of bacteria (data not shown), making it difficult to use in the preparation of gIII M13KO7-containing competent cells to take up the phagemid library.

In order to avoid these problems we attempted to simplify the system as much as possible by using a single genetic package, incorporating the genes for both interaction partners in the same phage vector. The circular genome of the filamentous phages M13, f1 and fd [25] consists of two transcription units, of 6 and 4 genes. The units are separated by a central terminator on one side and the origin of replication on the other (Fig. 3), and within the units there is no space between the open reading frames.

We inserted an antibiotic resistance gene into the fd genome to facilitate detection of infection events because plaque formation will occur only at near wt levels of replication and infectivity. Since initial experiments using fd-tet phage [26], which carries a tetracyline resistance gene in one of the hairpins of the phage origin of replication, yielded rather low phage titers, we decided to insert the resistance gene immediately downstream from the central terminator (Fig. 3b). In the wt phage, the −35 box of the weak gIII promoter is within the end of gVIII and the −10 box is within the strong central terminator.
(fd^{imm}). Therefore, we duplicated the -35 promoter box in gVIII and inserted resistance cassettes between the duplicated areas (Fig. 3b). We tested several antibiotic resistances at this position and found that the phages remained highly infectious but showed different titers (Table 1) and plaque morphologies with different resistances. To optimise the expression level of the resistance gene, we randomised its -35 promoter sequence. Any sequence isolated from a plaque and then from bacterial colonies was reasoned to lead to a level of expression sufficiently low to be compatible with phage propagation but showing different titers (Fig. 3c). In this orientation, however, homologous recombination occurred at a very high frequency between the duplicated glycine-rich regions, leading to restoration of a wt-like gIII.

In the next version, therefore, we reversed the orientation of the gIII partners by placing scFv-CT upstream of N1-N2-Ag (Fig. 3d). The two fusions were expressed under the control of separate lac promoters (lac^{imm}). Following removal of the original gIII, the expression cassette was inserted directly after the cam^{R} and the newly-inserted hp terminator (hp^{erm}) [27], and before the gIII promoter (gIIIp) and fd^{imm}. Infectious phages with wt properties were again isolated. In this case wt gIII was restored by intermolecular recombination, leading to duplication of genes between the glycine-rich regions (Fig. 3d).

In the third version of the system, we constructed a phage that does not rearrange at noticeable frequencies by almost completely eliminating the glycine-rich stretches. In the case of the scFv-CT, gIII was cut down to aa 250 and in case of N1-N2-Ag, to aa 219 (Fig. 3e).

As a model system, we have used the well-characterised antibody 179 which recognises the peptide epitope DVPDYA (hag) from hemagglutinin [16]. We tested the system by co-expressing the cognate anti-HAG scFv 179 or the non-cognate anti-HER2 scFv 4D5 [24,28] with the N1-N2-hag peptide fusion. We chose the antibody 4D5 as a control because we have found it to be extremely well expressed, and thus it should be displayed at a high level. Any non-specific positive effect of scFv-CT on the infectivity of its phage particle will therefore favour the growth of the phage displaying the 4D5 antibody. Similar numbers of phages (10^10-10^11) carrying cognate or non-cognate pairs of fusion proteins were used to infect cells. Of the 10^11 phage particles displaying anti-HAG scFv together with N1-N2-hag, we obtained 10^10 and non-resistant colonies for the non-cognate anti-HER2 scFv (Table 2). Restriction analysis of twelve colonies gave rise to the expected pattern, indicating that no rearrangement had occurred. We conclude that a very selective infection does indeed occur by the cognate antibody-antigen interaction, mediating the docking of probably several N1-N2-hag proteins to the multivalent antibody-displaying phage. As a further control we deleted the epitope from N1-N2-Ag and, again, no infectivity was observed in 10^11 phage particles. In a competitive experiment, only phages displaying anti-HAG scFv were isolated from a 1:100 mixture of anti-HAG to anti-HER2-carrying phages that co-displayed N1-N2-hag. Finally, under the conditions described, no background is observed (Table 3).

Ongoing experiments explore the use of this system to randomise the epitope of 179 as well as to map permissible changes in the antibody. In this way we will be able to evaluate the limits of on-rate, off-rate and binding constant permissible for in vivo SIP. For non-peptidic antigens, we have also purified the different N-terminal domains of gIIIp and subsequently chemically coupled these to several haptons. This will allow a more quantitative description of the infective process.
with respect to the N1-N2-Ag and N1-Ag fusions (C. Krebber et al., unpublished).

Only those cells in which a cognate interaction mediates the binding of the two fusion proteins give rise to infectious phages. Since the phage genome carries the information for both the antigen and the antibody, selection can be carried out from a pool of either fusion protein. If pools of both partners are present, cognate pairs will be selected cooperatively in a co-evolutionary fashion. We believe that this system will be particularly useful for simultaneously selecting antibodies against members of antigen libraries, e.g. cDNA libraries. Furthermore, because of the simplicity of the system, mutagenesis of either partner can be carried through many generations. We foresee that the development of a continuous system will be achieved upon increasing the efficiency of the infection process through its mechanistic dissection.
Acknowledgements: We would like to thank Dr. Stefan Klauser, Anke Kreberger, Dr. Gerard Wall, Dr. Kosi Gramatikoff, Prof. Walter Schaffner (Universität Zürich), Dr. Vic Ilag, Dr. Liming Ge and Dr. Simon Moroney (Morphosys GmbH München), for technical help and discussions and Morphosys GmbH for financial support.

References